The Role of Sorghum Legume Intercroping System in Improving Soil Productivity on Small Holder Farmers in Western Kenya

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Abstract: **Declining crops yield in the smallholder farmers cropping systems of sub-Saharan African (SSA) present the need to develop more sustainable production systems. Depletion of essential plant nutrients from the soils have been cited as the main contributing factors due to continues cultivation of cereal crops without application of organic/ inorganic fertilizers. Field experiments to evaluate effect of phosphorus (P) fertilizers, organic and integration of legumes in sorghum cropping systems on soil, available nitrogen (N) and P, were conducted in Busia County of Kenya during the short (SRS) and long rain seasons (LRS) of 2016 and LRS of 2017 respectively. The experiments comprised either soybean, common bean groundnut or sesame grown with sorghum. The design was a split plot in a randomized complete block design. Main plots were fertilizer inputs; Mavuno, FYM or their combination. Subplots comprised of the legume intercrops mentioned above. Application of Mavuno, FYM or their combination resulted in significantly higher legume, sesame crop yields above the control in the second season. Legume crops due to their N-fixation, litter fall and mineralization made availability of P and N. possible. Application of Mavuno, FYM or their combination gave comparable results with respect to the intercrop yields. Since FYM and (Mavuno+FYM) is cheaper than Mavuno, growing either soybean, common bean groundnut or sesame intercropping system with sorghum with application of the above is recommended for improved legume grain yields and soil fertility improvement.**

Keywords: **organic/inorganic inputs, legume cropping system, biomass decomposition, grain yields**

I. INTRODUCTION

In Kenya, challenges of food security, poverty and income inequalities remain a major concern for the Government despite policies on self-sufficiency in food being emphasized (Ombaka *et al,* 2014; Lokuruka, 2020).Continuous monocropping of maize without crop diversification on small pieces of the land with little or no provision for soil fertility maintenance contribute to the rapid depletion of soil nutrients in general and nitrogen in particular (Girijesh *et al.,* 2017; Grote, · 2021).

Attaining optimum crop yields in smallholder farms of Western Kenya remains a predicament with most farmers recording low annual harvests (Mwaura, 2021). This situation further translates into food insecurity and poverty especially in a country like Kenya where majority (> 70 %)of its population depends directly or indirectly on agriculture-related farm and off-farm activities for their livelihoods (Ng'ang'a, *et al.,* 2017). A report by FAO (2018) indicated that 60 percent of the population was living below the 1 dollar-a-day poverty line. Given that agriculture is a major contributor to the country's Gross Domestic Product (GDP) and revenue, this alarming trend is worrying and calls for urgent response in terms of agricultural policies (FAO, 2018).

The causes of these low crop yields are diverse with factors such as declining soil fertility taking the lead position (Vanlauwe, *et al.,* 2008). Soil infertility in Western Kenya smallholder farms is further escalated by various factors such as lack of /or inadequate use of inorganic/organic fertilizers, among other factors (Wawire,2021).In order to address the problem of soil infertility, the Kenyan government through the National Accelerated Agricultural Inputs Access Programme (NAAIAP) introduced subsidized fertilizers (GOK, 2014). This was aimed at raising fertilizer use to optimal levels in order to increase crop productivity from increased input use thereby raising land and labour productivity and food security for small holder farmers who form majority of households in Western Kenya (Ochola, & Fengying, 2015; Birch, 2018). Despite these efforts no much crop yields have been achieved even after devoting much land to the targeted crops under the fertilizer subsidy by 15 percent (Druilhe, & Barreiro-Hurlé, 2012; Lencucha *et al.,* 2020).

Soil fertility management approaches play a leading role in ensuring sustainable crop production on low nutrient soils (Urmi, 2022) such as those in western Kenya region. Increasing human population and the associated increased demand for food production on the other hand and food quality in the world require that proposed agronomic strategies for improvement should, in general, avoid high input costs associated with crop input costs. Organic inputs such as farm yard manure (FYM) and non-acidic fertilizer inputs like Mavuno are steadily gaining increased popularity and recognition from scientists as a means to improving soil productivity (Sharma, 2022). This could be due to the fact that they pose no ecological threats, have a longer-lasting effect on the soil, and, if well managed, they can often out yield recommended doses of chemical fertilizers (Mahdi and Mustafa, 2005; Hlisnikovský, 2022).

Legumes form a major component of every farming systems in SSA, making positive contributions to improving soil fertility and food security (Amede, 2003; Yuvaraj,2020). Legumes are potential sources of plant nutrients that complement/supplement inorganic fertilizers for cereal crops because of their ability to fix biological nitrogen (N) when included to the cropping systems ( ). By fixing atmospheric N_2, legumes offer the most effective way of increasing the productivity of poor soils either in monoculture, intercropping, crop rotations, or mixed cropping systems (Befekadu *et al.,* 2018). The complimentary nature of nitrogen fixing legumes also results in higher crop yields (Fan *et al.*,2006; Karavidas,2022;) besides ensuring economic utilization of land, labour and capital (Jeyabal and Kuppuswamy, 2001; World Bank, 2020).The information regarding optimum legume crop yields and associated soil fertility benefits is invariably unavailable for various agro-ecosystems in Sub-Saharan Africa( ). The present study aims at offering an answer as to whether legume crop diversification through different legume species and other high value traditional crops such as sesame under organic and inorganic inputs can result into improved soil fertility and sustainable increased crop productivity. The information obtained from the study can be used to develop interventions that may eventually result in increasing soil fertility, improving food security and/or improved livelihood for smallholder farmers who are the major players in cultivation of these crops from their farm production. The study considered the challenges the rural people face in farming which is their primary economic activity and tries to come up with innovations of optimizing their production from their small sized pieces of land.

II. MATERIALS AND METHODS

Study areas

Table 1. Characteristics of experimental sites.

Study site	Soil Type	Altitude	Rainfall	Mean Temperature	Latitude	Longitude
Busia Agricultural Training Centre	Orthic Ferralsols	1130-1375 m.a.s.l	1270-1790	14-30°C	0^0 16’45 N,	$34^0$20’ 20 E
Teso South (Farmer’s Field)	Gleyic Acrisols	1100-1400 m.a.s.l	1270-1790 mm	26-33 °C	0^0 33’40 N	34^0 31’ 06 E

Source: Jaetzold, et. al,. (2011)

Experimental design

The experiment was laid out in a split plot arrangement in RCBD design in 3 replications in each site and season. The main plots comprised of the fertilizer materials i.e., Mavuno fertilizer, Manure, Mavuno fertilizer+Manure and the Control. The subplots were formed by the different intercrops

Treatment application

The treatments were applied once at the start of the first season, 2016 in Busia and Teso at their recommended rates (Table 1).The treatments were replicated three times for each site and applied in plots measuring 5x4.5 m giving plot area of $22.5m^2$.In the first season (September –December 2016) the crop received the recommended fertilizer levels (band applied) as a blanket inputs under short rains as described in table 1 above. The phosphorus contribution from each input treatment was maintained at 26kg P $ha^{-1}$for this study, this rate being the recommended P level for most of the cereal crops within the western Kenya region for their optimum performance. The legumes and sesame intercrop were supplied with 30kg P. ha^{-1} according to.(Anochili, 1984)

Planting of field experiment

Field experiments commenced during the 2016 short rain season (SRs) (August to December 2016) and replicated during the 2017 long rains (LRs) (March to May 2017). Seredo Sorghum variety from Kenya Seed Company was drilled at seed rate of 3 kg ha^{-1}also at the pacing of 0.75 x 0.2 m. The intercrops i.e. common bean and soybean seeds were sourced from KALRO Kakamega and sesame and ground nut seeds were bought from the local markets in western Kenya. These were planted in between the sorghum rows at the recommended spacing of 0.33 from sorghum row x 0.15m within the row. The crop management practices i.e. gapping, weed control and top dressing with nitrogenous fertilizer, were done accordingly.

2.1. Soil Sampling and preparation

Initial composite soil samples were collected from a depth of 0–20 cm before planting the Test legume crops in 2016 short rains. Soil samples were also taken from each plot at planting and after each crop was harvested to assess the changes in soil fertility status due to the effect of legume precursor crop and integrated application of organic and inorganic fertilizers.

Soil pH was measured using Glass electrode method (H_2O) meter. The Walkley–Black method was followed for the determination of soil organic carbon (%OC), whereas the cation exchange capacity (CEC), in centimol (cmol)/100 g soil) was analyzed using ammonium acetate. Total nitrogen (%N) and available phosphorus (mg kg^{-1} soil) were analyzed by the Kjeldahl and the Olsen methods, respectively. The procedures for the methods used are outlined in Okalebo *et al.,* 2002. The result of initial soil analysis showed that the pH was in the strongly acidic range as per the standard classification procedure by Motsara and Roy (2008). Total nitrogen of the sites was highly variable and found in the low to moderate range. The available phosphorus content of the soil was categorized under very low range for both sites. Soil nitrogen and the organic carbon content varied in each site, though it was in the medium range for both sites

Table 2:Physico chemical properties of soil at 0-20 cm depth.

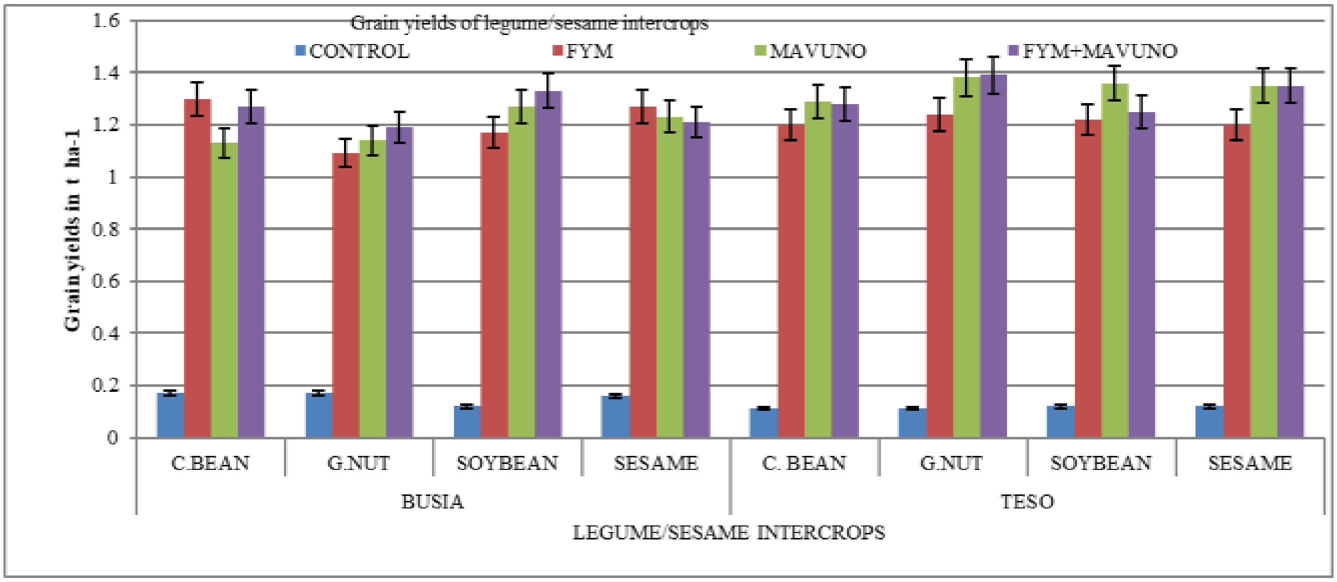
		(mg kg$^{-1)}$	(%)			(cmolkg^{-1}					(%)				
SITE	pH	P.	N	OC	C:N	Ca	Mg	K	Na	Al^{3+}+H^{+}	Sand	Silt	Clay	Text class	Order
Busia	4.85	1.82	0.21	1.50	7:01	0.90	0.35	0.37	0.96	1.95	55	24	21	Loam	Orthic Ferralsols
Teso	5.13	1.85	0.17	1.54	9:01	0.97	0.46	1.02	0.97	1.86	78	10	12	Sandy Loam	Gleyic Acrisols

III. RESULTS

1.Residual effects of treatment on legume and sesame grain yields

The data on legume intercrop and sesame grain yields are presented in Figure 1.

Figure 1: Effect of treatments on legume intercrop and sesame grain yields (t ha^{-1}) at Busia and Teso during the 2017 LR cropping season.



Of importance to note here is that none of the legume and sesame intercrops planted during the 2016 SR season reached harvesting stage due to intense moisture deficit experienced during this period. However, data for yields obtained from the test intercrops (common beans, groundnut, soybean and sesame) in 2017 LR due to the application of the soil fertility improvement materials during the 2016 SR are illustrated in Figure 1 for Busia and Teso sites. As observed, all the soil inputs gave significantly *(p<0.05)* higher crop yields above the control.

Fertilizer material application significantly (p<0.05) contributed to increased legume and sesame grains above the control at both sites Busia and Teso. In general crop yields were improved when the different nutrient materials were applied irrespective of the nutrient sources. For each of the materials applied, the results could be explained according to the findings of Cooke (1967) and Cui, 2021) who observed that the residual fertilizers left in the soil often raise crop yields that are hard to imitate with fresh fertilizer applications. Soil pH is especially important in maintaining fertilizer nutrient in the available forms. Hence the availability of Ca, phosphorus from *Mavuno* (Table 1) fertilizer applied, moisture supply from the rains during growth period and the possible release of other nutrients from organic matter mineralization with improved uptake that could have contributed to higher yields in the residual experiment. According to (Zerihun, *et al*., 2013, Ghosh,2022), organic manure alone or in combination with inorganic manure supplies nutrients throughout the season for the growth and development of the crop, in addition to improving soil moisture holding capacity and, thus, leading to increased productivity of the test crops. According to (Barber *et al.,*2014; Mahmud,2022), one of the benefits of applying soil amendments in the soil is to increase available molybdenum which is the main factor that enhances growth of legume crops on acid soils such as those in Busia and Teso study sites. This could partly explain the high yields obtained from the material amended plots in this study. Low soil nitrogen and phosphorus on the other hand are among the major factors limiting production and productivity of legume crops (Salama, 2021). Legume plants that depend on biological N_2 fixation for their N supply like soybean and others in this study require more P than plants receiving fertilizer N since the reduction of atmospheric N by the nitrogenous system is a very energy-consuming process (Stagnari, 2017). Hence, the plants require more P and other nutrients for symbiotic N fixation than for general plant metabolism (Kamara, Kwari, Ekeleme, Omoigui, & Abaidoo, 2008; Schelze, Temple, Beschow, & Vance, 2006; Míguez-Montero, 2020). Nitrogen is the most important nutrient for crop production, and its deficiency occurs in most countries of the world, Kenya included (Schelze, *et al.,* 2006, Kihara,2020). Evidence in the present study reveals the superiority of organic fertilizers in combination with inorganic sources and integration of legume crops in enhancing nutrient availability, optimizing soil environment that contribute to improved crop productivity.

2. Effect of cropping systems on % Soil Nitrogen at Busia and Teso

The results on the effect of intercropping system on % soil nitrogen content are presented in Table 3.

TABLE 3: Effect of Cropping Systems on % soil Nitrogen At Busia and Teso Over the study Period.

	........................DAYS FROM 1^{ST} PLANTING..................							
	BUSIA				**TESO**			
CROPPING SYSTEM	**60**	**120**	**180**	**240**	**60**	**120**	**180**	**240**
S- MONOCROP	0.22a	0.29ab	0.22a	0.27a	0.21ab	0.22 a	0.21ab	0.18a
S+GROUND NUT	0.24ab	0.32b	0.25ab	0.28a	0.21ab	0.22a	0.19a	0.16a
S+SESAM	0.24ab	0.29ab	0.21a	0.29a	0.19 a	0.25 a	0.24 b	0.18a
S+SOYBEAN	0.34b	0.28ab	0.26 b	0.29a	0.24 b	0.23 a	0.24b	0.19a
S+COMMON BEAN	0.26ab	0.27 a	0.24ab	0.27a	0.21a	0.25a	0.22ab	0.18a
MEANS	0.26	0.29	0.23	0.28	0.21	0.23	0.22	0.18
LSD$_{(0.05)}$)	0.11	0.05	0.04	ns	0.11	0.05	0.04	ns
CV	25.3	19.5	14.2	8.8	15.8	21.1	26.5	19.5

Means with the same alphabetical letter within a column are not significantly different at 5% probability using Fishers unprotected LSD value.

Ns- Treatment effects not significant according to Fisher's protected LSD

All the legume intercrops planted (common beans, sesame, soybean and common bean) with sorghum significantly ($p < 0.05$) contributed to the total soil nitrogen content for the Busia site in comparison with sorghum mono-crop plots. Sorghum-soybean intercropped plots recorded the highest N. levels of 0.34% N of soils sampled at 60 days from the planting of the first crop. Next was sorghum-common bean intercrop with 0.26%N, then 0.24 % N from the sorghum-common bean and sorghum sesame intercrops respectively. Finally, was control that recorded 0.22%N. General N% increases were observed across all treatments including the sorghum monocrop from soils sampled at 120 days (Table 3). The highest N values were observed with the sorghum-common bean intercrop when compared with the other sorghum intercrop systems. General declines in N% were observed at 180 days across all the intercrops. The highest N% contents were however realized at 240 days. The mean total soil N fluctuated for all treatments across the two cropping seasons. Teso site however recorded lower total N values for all the intercrops as compared to Busia site (Table 3).At (120 days from planting of the first crop) sorghum-soybean intercrop recorded the highest soil total nitrogen content. General N% reductions were observed at 180 and 240 days at Teso site. The fluctuation in soil N content in two sites could be explained in terms of crop nutrient uptake, immobilization by the soil microbes, leaching and volatilization due erratic weather conditions of high rainfall and temperature levels within western Kenya region. The low and declining N levels in Teso as compared to Busia site could be due to their difference in soil texture where soils in Teso had higher sand contents observed (Table 2) which could have encouraged more N leaching as opposed to the soils in Busia site leading to low soil nitrogen levels observed.

The increase in N availability in intercrops hosting legumes occurs because the competition for soil N from legumes is weaker than from other plants. Moreover, non-legumes obtain additional N from that released by legumes into the soil (Pappa *et al.,* 2012, White *et al.,* 2013) or via mycorrhizal fungi (Chen, 2022). Legumes can contribute up to 15% of the N in an intercropped cereal (LI *et al.,* 2009), thus increasing biomass production and carry-over effects (Pappa *et al.,* 2012), reducing synthetic mineral N-fertilizer use and mitigating N_2O fluxes (Beaudette, 2016).

3. Effect of cropping systems on Soil available P. at Busia and Teso

Table 4: effect of cropping systems on soil available phosphorus at Busia and Teso over the study period

	........................DAYS FROM 1^{ST} PLANTING..................							
	BUSIA				**TESO**			
CROPPING SYSTEM	**60**	**120**	**180**	**240**	**60**	**120**	**180**	**240**
S- MONOCROP	5.94a	6.58a	35.04a	40.08a	7.361a	2.05a	41.24ab	49.88a
S+GROUND NUT	6.34ab	6.71a	36.8a	42.36a	9.26a	1.84a	37.68ab	51.18a
S+SESAM	6.71ab	8.28a	31.3a	39.72a	9.13a	1.72a	35.8a	58.7a
S+SOYBEAN	7.59bc	7.19a	39.06a	45.57a	7.31a	1.96a	36.32b	54.43a
S+COMMON BEAN	8.72c	7.84a	29.76a	39.27a	7.68a	2.03a	49.22b	60.32a
MEANS	7.06	7.34	34.4	41.4	8.15	1.92	40.0	54.9
$LSD_{(0.05)}$	1.31	ns	ns	ns	ns	ns	11.7	16.47
CV	22.6	34.9	11.3	14.4	38.6	7.5	22.9	34.7

Means with the same alphabetical letter within a column are not significantly different at 5% probability using Fishers unprotected LSD value.

Ns- Treatment effects not significant according to Fisher's protected LSD.

Legumes intercropped with cereals can provide not only nitrogen, but also other minerals, soil cover, as they also smother weeds, provide habitat for pest predators, and increase microbial diversity, such as vesicular arbuscular mycorrhizae (VAM). VAM, a fungus, plays an interesting role in that it is thought to facilitate nutrient transfer e.g., phosphorus to the other crop. The association with VAM becomes very significant where one crop has the ability to mine different sources of nutrients than the other. Some evidence shows more P, K, Ca, and Mg availability in intercrops than in monocultures (Vandermeer 1992; Li *et al.* 2007; Begum, 2019).

As observed in Table 4 all the intercrops (common beans, sesame, soybean and groundnut) planted with sorghum significantly ($p < 0.05$) contributed to the total soil available P content for the Busia site when compared to the sorghum mono-crop plots. The highest P Contents were observed in the sorghum-common bean intercropped plots which recorded 8.72 (mgkg^{-1}) from the soils sampled at 60 days from the planting of the first crop. This was followed by the sorghum-soybean intercrop with 7.59mgkg^{-1}P.Next was 6.71 mgkg^{-1}**P** which was observed from the sorghum- sesame intercrops. The least P contents were recorded in sorghum-common bean and control plots with the P mean values of 6.34 and 5.94 mgkg^{-1} P respectively. General increases in P values were observed across all intercrops including the sorghum mono-crop except in the sorghum- soy bean intercrop which recorded a decrease in the soil available P content when the soils were sampled at 120 days (Table 4) at Busia site. At 180 days all the intercrop treatments showed an upward trend in available soil P contents. These improvements in available P contents were realized at 240 days from the start of the study period. The mean total soil P followed the same increasing trend for all treatments across the two cropping seasons.

On the contrary, Teso site generally recorded higher available soil P values for all the intercrops as compared to Busia site except for the sorghum-soybean and sorghum-common bean intercrops (Table 4).Similar to Busia site, general reductions in P values were observed at 120 days across all the intercrops, including the sorghum mono-crop. Highest values were still realized at180 and 240 days from the start of the study period which showed an increase in soil available P. Plant nutrients availability may be influenced by some plant growth regulators through synthesizing plant hormones or facilitating the uptake of nutrients from the soil by different direct mechanisms, e.g., atmospheric nitrogen (N) fixation, solubilization of phosphorus (P),and synthesis of siderophores for iron sequestration, making nutrients available to plants,(Glick, *et al.*, 2007;Jaiswal, 2021;). According to (Askegaard & Eriksen, 2008 and Giordano,2021), legumes as a catch crop is able to reduce nitrate and K leaching and act not only as a N_2 fixing crop but also as a catch crop by taking up additional soil minerals N,P, and K . (Flores-Sanchez *et al*., 2013). These nutrients can easily be released in the soil upon litter fall and decomposition which can readily be taken by the crop from the soil solution by the subsequent crop (Giweta, 2020) These findings make legumes an important tool in the cropping systems where N, P and K are the major yield limiting factors such as those found in western Kenya region. The higher P levels observed in the sorghum mono and intercropped plots especially at the later stages of sampling in this study could be due to organic acid mineralization of the soil nutrients from the crops litter fall and decomposition and their easy dissolution in soil solution to release the contained nutrients including the P observed in this study. When plants are subjected to low P soil conditions such as those in Busia and Teso (Table 2), according to (Gilbert *et al.,* 1999; Richardson *et al.,* 2001 and Dixon, 2020), secretion of acid phosphatase from roots is a common response. For example, under such low P soil conditions, white lupin secretes an acid phosphatase capable of phytate degradation into the rhizosphere (Gilbert *et al.,* 1999; Dixon, 2020). P acquisition has also been improved through approaches aimed at increasing citrate synthesis in and/or exudation from plant cells. This approach is based upon the enormous evidence showing that exudation of citrate and malate from roots effectively solubilizes unavailable P sources (Marschner *et al*., 1995; Elhaissoufi, 2021).

In intercropping cowpea–maize, Latati *et al*., (2016) and Tang, 2021, found an increase in P availability at rhizosphere level associated with significant acidification than in sole cropping. Wang *et al.* (2012 and Hallama,2019), in their work related to N and P cycling in the rhizosphere of wheat and grain legumes (faba bean and white lupin) grown in monoculture or in wheat/legume mixtures, found that the less-labile organic P pools (i.e. NaOH-extractable P pools and acid-extractable P pools) significantly accumulated in the rhizosphere of legumes. However, the P uptake and the changes in rhizosphere soil P pools seem to depend also on legume species. Compared with the unplanted soil, the depletion of labile P pools (resin P and $NaHCO_3$-P inorganic) was the greatest in the rhizosphere of faba bean (54 and 39%) with respect to chickpea, white lupin, yellow lupin and narrow-leafed lupin (Hassan *et al.,* 2012). Of the less-labile P pools, NaOH-P inorganic was depleted in the rhizosphere of faba bean, while NaOH-P organic and residual P were most strongly depleted in the rhizosphere of white lupin (Hassan *et al*.,2012).

Some grain legumes, including chickpea, pigeon pea and white lupin can mobilize fixed forms of soil P through the secretion of organic acids such as citrate and malate and other P mobilizing compounds from their roots (Hocking, 2001;Homulle, 2021). Among grain legumes, white lupin most strongly solubilize P, a function that can be facilitated by its proteoid roots that may englobe small portions of soil (Angus, 2015; Pueyo, 2021). Glasshouse experiments using a highly P-fixing soil showed better wheat growth following white lupin than soybean (Hocking and Rindall, 2001), suggesting that the cereal was able to access P made available by the previous white lupin break crop.

Rhizosphere acidification by exudates leads to desorption of PO_4 from the soil matrix with a concomitant increase in P availability. Degraded and infertile soils such as those in western Kenya are realized as a result of continuous monocropping and insufficient organic matter reprocessing coupled with occurrence of rainfall variability marked by common dry spells account for low crop yield (Amos *et al*., 2012; Amwata, 2020). It was further noted that the understanding of the fact that maintenance and improvement of soil fertility cannot be exclusively through the use of predictable fertilizers (Amos *et al.,* 2012; Mucheru-Muna, 2021). As a trait in legumes as cover crops, conservation involves minimum soil disturbance, permanent soil cover with living or dead plant resources, and diversified `cereal cropping system associated by legumes crops (Amos *et al*.,2012; Kocira *et al*.,2020). The difference in soil P content in two sites could be explained in terms of crop nutrient uptake, immobilization by the soil microbes, leaching and P-sorption. The higher P levels in Teso as compared to Busia site could be due to their difference in soil texture and cropping duration. Soils in Teso besides being high in sand contents (Table 2) they were also under some period of fallowing implying higher initial inherent nutrient contents which could have contributed to more P as opposed to the soils in Busia site which are under constant cropping, being a farmers training centre. The situation which leads to soil nutrient depletion due to crop uptake (Swoboda, 2022).

IV. CONCLUSION

Major advantages of legumes include the amount of nitrogen fixed into the soil and the high quality of the organic matter released to the soil in term of C/N ratio. Some legume species have also deep root systems, which facilitate nutrients solubilization by root exudates and their uptake/recycling as well as water infiltration in deeper soil layers.

Legumes that can recover unavailable forms of soil phosphorus could be major assets in future cropping systems. Consequently, those legumes which are able to accumulate phosphorus from forms normally unavailable need to be further studied, since phosphorus represents an expensive and limiting resource in several cropping systems.

REFFERENCES

[1] Amos, R. N., Jens, B. A., & Symon, M. (2012). On farm evaluation of yield and economic benefits of short term maize legume intercropping systems under conservation Agriculture in Malawi.

[2] Amwata DA. (2020). Situational analysis study for the agriculture sector in Kenya. CCAFS Report. Wageningen, the Netherlands: CGIAR Research Program on Climate Change, Agriculture and Food Security (CCAFS).

[3] Angus JF, Kirkegaard JA, Hunt JR, Ryan MH, Ohlander L, Peoples MB. (2015). Break crops and rotations for wheat. Crop Pasture Sci.;66: 523–52.CrossRefGoogle Scholar

[4] Beaudette C, Bradley RL, Whalen JK, McVetty PBE, Vessey K, Smith DL.(2010). Tree-based intercropping does not compromise canola (Brassica napus L.) seed oil yield and reduces soil nitrous oxide emissions. Agric Ecosyst Environ.;139:339.CrossRefGoogle Scholar.

[5] Begum Naheeda, Qin Cheng, Ahanger Muhammad Abass, Raza Sajjad, Khan Muhammad Ishfaq, Ashraf Muhammad, Ahmed Nadeem, Zhang Lixin(2019). Role of Arbuscular Mycorrhizal Fungi in Plant Growth Regulation: Implications in Abiotic Stress Tolerance;Frontiers in Plant Science,Volume=10,PAGES=1068 URL=https://www.frontiersin.org/article/10.3389/fpls.2019.01068 Doi=10.3389/fpls.2019.01068,ISSN=1664-462X

[6] Birch, I. (2018). Agricultural productivity in Kenya: barriers and opportunities. K4D Helpdesk Report. Brighton, UK: Institute of Development Studies.

[7] Chen, David Kleijn, Jeroen Scheper, Thijs P.M. Fijen,(2022).Additive and synergistic effects of arbuscular mycorrhizal fungi, insect pollination and nutrient availability in a perennial fruit crop, Agriculture, Ecosystems & Environment,Volume 325,107742,ISSN 0167-8809,https://doi.org/10.1016/j.agee.2021.107742.(https://www.sciencedirect.com/science/article/pii/S0167880921004461))

[8] Dixon, Mary, Eric Simonne, Thomas Obreza, and Guodong Liu. (2020). "Crop Response to Low Phosphorus Bioavailability with a Focus on Tomato" Agronomy 10, no. 5: 617. https://doi.org/10.3390/agronomy10050617.

[9] Druilhe, Z., & Barreiro-Hurlé, J. (2012). Fertilizer subsidies in sub-Saharan Africa (No. 12-04). ESA Working paper.

[10] Egbe O., and Idoko J.1 (2012)."Evaluation of pigeonpea genotypes for intercropping with maize and sorghum in southern Guinea Savanna: Economic benefits." International journal of Agriculture and forestry 2. 108-114.

[11] Elhaissoufi Wissal, Cherki Ghoulam, Abdellatif Barakat, Youssef Zeroual, Adnane Bargaz, (2021).Phosphate bacterial solubilization: A key rhizosphere driving force enabling higher P use efficiency and crop productivity, Journal of Advanced Research,ISSN 2090-1232, https://doi.org/10.1016/j.jare.2021.08.014.(https://www.sciencedirect.com/science/article/pii/S2090123221001673)

[12] Food Security Report (Prepared by Kenya Agricultural Research Institute | Food Security Portal.(2018). Food securityportal.org. Retrieved 9 April 2018, from http://www.foodsecurityportal.org/kenya/food-security-report-prepared-kenya-agricultural-research-institute.

[13] Ghosh, D., Brahmachari, K., Skalický, M., Roy, D., Das, A., Sarkar, S., Moulick, D., Brestič, M., Hejnak, V.,Vachova, P., Hassan, M. M., & Hossain, A. (2022). The combination of organic and inorganic fertilizers influence the weed growth, productivity and soil fertility of monsoon rice. PloS one, 17(1), e0262586. https://doi.org/10.1371/journal.pone.0262586

[14] Gilbert GA, Knight JD, Vance CP, Allan DL (1999). Acid phosphatase activity in phosphorus-deficient white lupin roots. Plant Cell Environ (22):801–810.

[15] Giordano, Maria, Spyridon A. Petropoulos, and Youssef Rouphael. 2021. "The Fate of Nitrogen from Soil to Plants: Influence of Agricultural Practices in Modern Agriculture" Agriculture 11, no. 10: 944. https://doi.org/10.3390/agriculture11100944.

[16] Giweta, M.(2020). Role of litter production and its decomposition, and factors affecting the processes in a tropical forest ecosystem: a review. j ecology environ **44,** 11. https://doi.org/10.1186/s41610-020- 0151-2

[17] Glick,B. R.,Todorovic, B., Czarny, J., Cheng, Z., Duan, J., & McConkey, B. (2007). Promotion of plant growth by bacterial ACC deaminase. Crit. Rev. Plant Sci .(26), 227-242.

[18] Grote U, Fasse A, Nguyen TT and Erenstein O (2021). Food Security and the Dynamics of Wheat and Maize Value Chains in Africa and Asia. Front. Sustain. Food Syst. 4:617009. doi: 10.3389/fsufs.2020.617009.

[19] Hallama, M., Pekrun, C., Lambers, H. et al.(2019). Hidden miners – the roles of cover crops and soil microorganisms in phosphorus cycling through agroecosystems. Plant Soil **434,** 7–45 (2019). https://doi.org/10.1007/s11104-018-3810-7

[20] Hassan HM, Hasbullah H, Marschner(2013). P. Growth and rhizosphere P pools of legume–wheat rotations at low P supply. Biol Fertil Soils.;49:41–9.CrossRefGoogle Scholar.

[21] Hlisnikovský, L.; Menšík,L.; ˇCermák, P.; Kˇrížová, K.; Kunzová,(2022).E. Long-Term Effect of Pig Slurry and Mineral Fertilizer Additions on Soil Nutrient Content, Field Pea Grain and Straw Yield under Winter Wheat–Spring Barley–Field Pea Crop Rotation on Cambisol and Luvisol. Land, 11, 187. https://doi.org/10.3390/land1102018.

[22] Hocking PJ, Randall PJ. (2001). Better growth and phosphorus nutrition of sorghum and wheat following organic acid secreting crops. In: Horst WJ, et al., editors. Proceedings of the 14th international plant nutrition colloquium Germany. Dordrecht: Kluwer Academic Publishers;. p. 548–9.Google Scholar.

[23] Homulle, Z., George, T.S. & Karley, A.J.(2022). Root traits with team benefits: understanding belowground interactions in intercropping systems. Plant Soil **471,** 1–26 (2022). https://doi.org/10.1007/s11104-021-05165-8

[24] Ibewiro, B.,Sanginga, N., Vanlauwe, B., & Merky, R. (2000) Nitrogen contribution from decomposing cover crop residues to maize in tropical derived savanna. Nutrient Cycling in Agroecosystems, 57, 131-140. http://dx.doi.org/10.1023/A:1009846203062.

[25] IUSS Working Group,W.(2006). World reference base for soil resource 2006.A frame work for international classification,correlation and communication. World soil Resource Report No.(103).FAO,Rome.

[26] Jaiswal Sanjay K., Mohammed Mustapha, Ibny Fadimata Y. I., Dakora Felix D.(2021). Rhizobia as a Source of Plant Growth-Promoting Molecules: Potential Applications and Possible Operational Mechanisms. Frontiers in Sustainable Food SystemsVol 4pp.311https://www.frontiersin.org/article/10.3389/fsufs.2020.619676 doi.org/10.3389/fsufs.2020.619676 -ISSN=2571-581X

[27] Jeyabal, A. and KuppuswamyG.(2001).Recycling of organic wastes for the production of vermicompost and its response in rice-legume cropping systems and soil fertility. Heriana J. Agron.,(15): 153–170.

[28] Kamara, A. Y., Kwari, J., Ekeleme, F., Omoigui, L., & Abaidoo, R. (2008). Effect of Phosphorus Application and Soybean Cultivar on Grain and Dry Matter Yield of Subsequent Maize in the Tropical Savannas of North-EasternNigeria. Afr. J. Biotechnol., , (7), 2593–2599.

[29] Karavidas, I.; Ntatsi, G.;Vougeleka, V.; Karkanis, A.; Ntanasi,T.; Saitanis, C.; Agathokleous, E.;

[30] Ropokis, A.; Sabatino, L.; Tran, F.;et al.(2022). Agronomic Practices to Increase the Yield and Quality of Common Bean (Phaseolus vulgaris L.): A Systematic Review. Agronomy,12, 271. https://doi.org/10.3390/agronomy12020271

[31] Kenya at a glance|FAO in Kenya|Food and Agriculture Organization of the United Nations. (2018). Fao.org. Retrieved 9 April 2018, from http://www.fao.org/kenya/fao-in-kenya/kenya-at-a-glance/en/

[32] Kocira, Anna, Mariola Staniak, Marzena Tomaszewska, Rafał Kornas, Jacek Cymerman, Katarzyna Panasiewicz, and Halina Lipińska. (2020). "Legume Cover Crops as One of the Elements of Strategic Weed Management and Soil Quality Improvement. A Review" Agriculture 10, no. 9: 394. https://doi.org/10.3390/agriculture10090394.

[33] Kihara, J., Bolo, P., Kinyua, M. et al.(2020). Micronutrient deficiencies in African soils and the human nutritional nexus: opportunities with staple crops. Environ Geochem Health 42, 3015–3033 (2020). https://doi.org/10.1007/s10653-019-00499-w

[34] Latati M, Bargaz A, Belarbi B, Lazali M, Benlahrech S, Tellah S.(2016). The intercropping common bean with maize improves the rhizobial efficiency, resource use and grain yield under low phosphorus availability. Eur J Agron.;72: 80–90.CrossRefGoogle Scholar.

[35] Lencucha, R., Pal, N.E., Appau, A. et al. (2020). Government policy and agricultural production: a scoping review to inform research and policy on healthy agricultural commodities. Global Health **16,** 11 https://doi.org/10.1186/s12992-020-0542-2

[36] Li L, Zhang L-Z, Zhang F-Z.(2013). Crop mixtures and the mechanisms of over yielding. In: Levin SA, editor. Encyclopedia of biodiversity, vol. 2. 2nd ed. Waltham: Academic Press;p. 382–95.CrossRefGoogle Scholar.

[37] Li YF, Ran W, Zhang RP, Sun SB, Xu GH (2009). Facilitated legume nodulation, phosphate uptake and nitrogen transfer by arbuscular inoculation in an upland rice and mung bean

intercropping system. Plant Soil.;315:285–96.CrossRefGoogle Scholar.

[38] Lokuruka, M. N. (2020). 'Food and Nutrition Security in East Africa (Kenya, Uganda and Tanzania): Status, Challenges and Prospects', in B. Mahmoud (ed.), Food Security in Africa, Intech Open, London. 10.5772/intechopen.95036.

[39] Mahmud MS, Chong KP. (2022).Effects of Liming on Soil Properties and Its Roles in Increasing the Productivity and Profitability of the Oil Palm Industry in Malaysia. Agriculture; 12(3):322. https://doi.org/10.3390/agriculture12030322

[40] Marschner H. (1995).Mineral Nutrition of Higher Plants, 2nd edition.Academic Press, Munchen.

[41] Motsara, M.L.; Roy, R.N.(2008). Guide to Laboratory Establishment for Plant and Nutrients Analysis; FAO Fertilizer and Plant Nutrition Bulletin 19; Food and Agriculture Organization of the United Nations: New Delhi, India.

[42] Mucheru-Muna Monicah Wanjiku , Mildred Achieng Ada, Jayne Njeri Mugwe, Franklin Somoni Mairura, Esther Mugi-Ngenga, Shammie Zingore, James Kinyua Mutegi,(2021). Socio-economic predictors, soil fertility knowledge domains and strategies for sustainable maize intensification in Embu County,Kenya,Heliyon,Volume 7, Issue 2, e06345, ISSN 2405-8440,https://doi.org/10.1016/j.heliyon.2021.e06345. (https://www.sciencedirect.com/science/article/pii/S2405844021004503)

[43] Mwaura George G., Kiboi Milka N., Bett Eric K., Mugwe Jayne N., Muriuki Anne, Nicolay Gian, Ngetich Felix K. (2021).Adoption Intensity of Selected Organic-Based Soil Fertility Management Technologies in the Central Highlands of Kenya.Frontiers in Sustainable Food Systems

[44] Ng'ang'a, Stanley Karanja, An Notenbaert, Chris Miyinzi Mwungu,Caroline Mwongera, and Evan Girvetz."(2017). "Cost and benefit analysis for climate-smart soil practices in Western Kenya.

[45] Ochola, R. O., & Fengying, N. I. E. (2015). Evaluating the effects of fertilizer subsidy programmes on vulnerable farmers in Kenya. Journal of Agricultural Extension and Rural Development, 7(6), 192-201.

[46] Ombaka DM. (2014). Of Kenya's eaters and eatists: Hunger as a development and social justice challenge. J Soc Welf Hum Righ. 2(1):107-129.

[47] Okalebo, J. R., Gathua, K. W. & Woomer, P. L. J. (2002). Laboratory Methods of Soil andPlant Analysis: A Working Manual (2nd Edition). Nairobi, Kenya. Marvel EPZ K Ltd.

[48] Pappa VA, Rees RM, Walker RL, Baddeley JA, Watson CA.(2012). Legumes intercropped with spring barley contribute to increased biomass production and carry-over effects. J Agric Sci.;150:584–94.CrossRefGoogle Scholar.

[49] Pueyo, J. J., Quiñones, M. A., Coba de la Peña, T., Fedorova, E. E., and Lucas, M. M. (2021).Nitrogen and phosphorus interplay in lupin root nodules and cluster roots. Front. Plant Sci. 12:644218. doi: 10.3389/fpls.2021.64421

[50] Richardson AE (2001). Prospects for using soil microorganisms to improve the acquisition of phosphorus by plants. Aust J Plant Physiol 28: 897–906.

[51] Urmi, T.A.; Rahman, M.M.;Islam, M.M.; Islam, M.A.; Jahan, N.A.; Mia, M.A.B.; Akhter, S.;Siddiqui, M.H.; Kalaji, H.M. (2022).Integrated Nutrient Management for Rice Yield, Soil Fertility, and Carbon Sequestration. Plants , 11, 138. https://doi.org/10.3390/plants 11010138.

[52] Salama, H., Nawar, A. I., Khalil, H. E., & Shaalan, A. M. (2021). Improvement of Maize Productivity and N Use Efficiency in a No-Tillage Irrigated Farming System: Effect of Cropping Sequence and Fertilization Management. Plants (Basel, Switzerland), 10 (7), 1459. https://doi.org/10.3390/plants10071459.

[53] Schelze, J., Temple, G., Temple, S. J., Beschow, H., & Vance, C. P. (2006). Nitrogen Fixation by White Lupin under Phosphorus Deficiency Nitrogen Fixation by White Lupin under Phosphorus Deficiency. Ann. Bot., (98), 731–740.

[54] Sharma SB (2022). Trend setting impacts of organic matter on soil physico-chemical properties in traditional vis -a- vis chemical-based amendment practices. PLOS Sustain Transform 1(3): e0000007. https://doi.org/10.1371/journal. pstr.0000007.

[55] Stagnari, F., Maggio, A., Galieni, A. et al. (2017). Multiple benefits of legumes for agriculture sustainability: an overview. Chem. Biol. Technol. Agric. **4,** 2 .https://doi.org/10.1186/s40538-016-0085-1.

[56] Swoboda Philipp, Thomas F. Döring, Martin Hamer,(2022).Remineralizing soils? The agricultural usage of silicate rock powders: A review,Science of The Total Environment,Volume 807, Part 3,150976,ISSN 0048 9697,https://doi.org/10.1016/j.scitotenv.2021.150976.(https://www.sciencedirect.com/science/article/pii/S004896972106054X)

[57] Tang, X., Zhang, C., Yu, Y. et al.(2021). Intercropping legumes and cereals increases phosphorus use efficiency; a meta-analysis. Plant Soil **460,** 89–104 (2021). https://doi.org/10.1007/s11104-020-04768-x.

[58] Vanlauwe, B., Kanampiu, F., Odhiambo, G. D., De Groote, H., Wadhams, L. J., & Khan, Z. R. (2008). Integrated management of Striga hermonthica, stem borers, and declining soil fertility in western Kenya. Field Crops Research, 107(2), 102-115.

[59] Vandermeer J. (1993). The ecology of Intercropping. Cambridge University Press.

[60] Walkley, A.; Black, C.A. An examination of Degtjareff method for determining soil organic matter and the proposed modification of the chromic acid titration method. Soil Sci. **1934**, 37, 29–38. [CrossRef]

[61] Wang Y, Marschner P, Zhang F. (2012). Phosphorus pools and other soil properties in the rhizosphere of wheat and legumes growing in three soils in monoculture or as a mixture of wheat and legume. Plant Soil.;354: 283–98.CrossRefGoogle Scholar.

[62] Wawire, A. W., Csorba, Á., Tóth, J. A., Michéli, E., Szalai, M., Mutuma, E., & Kovács, E. (2021). Soil fertility management among smallholder farmers in Mount Kenya East region. Heliyon, 7(3), e06488. https://doi.org/10.1016/j.heliyon.2021.e06488.

[63] World Bank (2020). Commodities markets outlook: implications of COVID-19 for commodities. April. Available from https://openknowledge.worldbank.org/bitstream/handle/10986/33624/CMO- April-2020.pdf

[64] Yuvaraj, M., Pandiyan, M., & Gayathri, P. (2020). Role of Legumes in Improving Soil Fertility Status. In (Ed.), Legume Crops - Prospects, Production and Uses. IntechOpen. https://doi.org/10.5772/intechopen.93247

[65] Zerihun, A., Sharma.,J. J.,Nigussie, D., & Fred, K. (2013). The Effect of Integrated Organic and Inorganic Fertilizer Rates on Performances of Soybean and Maize Component Crops of a Soybean/maize Mixture.